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# Biocontrol of Fusarium Wilt in Tomato: An Eco-friendly and Cost Effective Approach

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ABSTRACT: Tomato is the second most widely produced vegetable in the world. All through growth time or even after harvesting, tomatoes are prone to several diseases brought about through viruses, fungal pathogen, bacteria, and nematodes. Tomato wilt is a fungal disease, caused by *Fusarium oxysporum* f.sp. *lycopersici*, that limiting tomato output severely around the world. Several measures were offered to restrict the spread of *Fusarium oxysporum* f.sp. *lycopersici* but were difficult because of the ability of the fungus to stay long in the soil. A wide range of chemical pesticides currently available and the continuous use of these pesticides affect the food substance of tomatoes as well as their texture and the performance of soil in order to control all these vulnerabilities. Thus, this review focuses on the possible use of microbial pesticide potential along with mechanisms for the management of wilt disease, nature of causal organism, epidemiology and losses, infection stages and disease cycle are also discussed. The use of a microbial pesticide is an environmentally friendly and effective way to prevent tomatoes from wilt disease and its devastating consequences. Different microorganisms have been used in tomato wilt treatment and are now being discussed. The beneficial inoculum not only suppresses the disease, but it also contributes in the healthy growth of crops.

Keywords: Microbial pesticides, Bioagents, PGPR, ISR, Trichoderma, Pseudomonas, Glomus, Baccillus

# INTRODUCTION

Tomato is a vital solanaceous crop grown worldwide significantly contributing in human nutrition. In tomato cultivation, the major concern is regarding Wilt caused by Fusarium oxysporum f.sp. lycopersici (Loganathan et al., 2009). An eco-friendly approach towards management of the disease will abolish the harsh outcome of chemicals like residual toxicity, pollution of environment, and development of resistance in pathogens against continuously used fungicide for their control. Enormous communities of bacterial residing in the rhizosphere and actively participating in growth promotion activity of plant are referred as plant growth promoting rhizobacteria (PGPR) (Kloepper et al., 1980). The PGPR creates competition for ecological niche/substrate by forming antibiotics, hydrogen cyanide, releasing siderophores and secretion of fungal cell wall lysis enzymes thus behave as biocontrol agents (Glick and Bashan 1997; Wang et al., 2000; Saravanakumar et al., 2007). PGPR additionally lead to activation of induced systemic resistance (ISR) in varied crops against several diseases (Kloepper and Beauchamp 1992; Liu et al., 1995; Chen et al., 2000; Sangeetha et al., 2010). For a wide range of crops, use of PGPR to control different pest and diseases and to improve production turned out be studied in depth, but knowledge regarding its effect on quality of fruit, especially about fruit texture and amount of lycopene available stays quite inadequate. Among different carotenoid, lycopene is one available in tomato which is a key composite required in supplement for humankind because it helps to reduce chances of prostate cancer diseases and lower down the cardiovascular and (Giovannucci prostate cancer diseases 1999: Giovannucci et al., 2002).

In the soil, other than PGPR, there are other microbes known as biocontrol agents like *Bacillus* spp., *Pseudomonas* spp., *Streptomyces* spp., *Trichoderma* spp. have been demonstrated to have a substantial effect against soil-borne pathogens that are extremely race specific (Upadhyay *et al.*, 2021) (Table 1).

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# Table 1: List of some biocontrol agents reported to act against Fusarium wilt disease of tomato caused by Fusarium oxysporum f.sp. lycopersici.

Sr. No.	<b>Biocontrol Agents</b>	Mechanisms	References
1	Bacillus subtilis	Induced the activities of defense	Loganathan et al., 2014
		related enzymes	
2	Pseudomonas	Increased resistance	M'piga <i>et al.</i> , 1997
	fluorescens		
3	Bacillus subtilis	Antagonistic activity	Mohammed and Toama 2019
4	Pseudomonas	Antagonistic activity	Mohammed and Toama 2019
	fluorescens		
5	Trichoderma harzianum	Antagonistic activity	Arenas et al., 2018
6	T. spirale	Antagonistic activity	Vargas-Inciarte <i>et al.</i> , 2019
7	Pseudomonas	ISR and antagonistic activities	Boukerma et al., 2017
	fluorescens		D 1 0015
8	Pseudomonas putida	ISR and antagonistic activities	Boukerma <i>et al.</i> , 2017
9	Trichoderma viride	Increase growth and chlorophyll	Wani and Mir 2009
10		content	Western 1 Min 2000
10	1. narzianum	Increase growth and chlorophyll	wani and Mir 2009
11	<u>Classica en esta e</u>		Wani and Min 2000
11	Giomus mossae	increase growth and chlorophyll	wani and Mir 2009
12	G fasciculatium	Increase growth and chlorophyll	Wani and Mir 2009
12	G. juscie ututtum	content	wall and will 2009
13	Bacillus sphaericus	Antagonistic activities	Wani and Mir 2009
14	Pseudomonas putida	Antagonistic activities	Kouki <i>et al.</i> 2012
15	Burkholderia gladioli	Antagonistic activities	Kouki <i>et al.</i> 2012
16	Stenotrophomonas	Inhibition of the systemic fungus	Avdi-Ben-Abdallah <i>et al</i> 2020
10	maltophilia	progress	riyar ben riodanan er an, 2020
17	Azotobacter	Inhibition of the systemic fungus	Avdi-Ben-Abdallah et al., 2020
	chroococcum	progress	
18	Serratia marcescens	Inhibition of the systemic fungus	Aydi-Ben-Abdallah et al., 2020
		progress	
19	Trichoderma virens	Induce JA and SA signalling	Jogaiah et al., 2018
		cascades for the elicitation of	-
		Fusarium oxysporum resistance	
20	Alcaligenes faecalis	Antifungal potential	Abdallah et al., 2016
21	Bacillus cereus	Antifungal potential	Abdallah et al., 2016
22	Bacillus	Anti-fungal secondary metabolites	Gowtham et al., 2016
	amyloliquefaciens	production	
23	Bacillus pumilis	Induction of resistance	Benhamou et al., 1998

Among them, in the worldwide the most explored are Trichoderma sp. (Howell and Stipanovic 1995); this is due to its ubiquity nature, its easy isolation property and its capacity to quickly grow on a huge number of substrates (Candela et al., 1995; Verma et al., 2007). Due to ongoing research for the past 10 years, isolation, selection and evaluation of native species of Trichoderma spp. have been done. The aim behind was to establish biological control against diverse diseases by anticipated numerous mechanisms and for the employment of this fungus providing satisfactory outcomes (Cook and Baker 1989; Chet et al., 1998; Sandoval 2011; Awad et al., 2014; Hamed et al., 2015). Trichoderma's mechanism of action with different plant pathognes may turn collaborative Septoria triticii, Sclerotium rolfsii, Sclerotiniasclerotiorum, Rhizoctonia solani, Fusarium oxysporum and Pythium splendens in wheat, cucumber, soybean and lettuce, soybean, tomato and beans, respectively (Ghisalbertí et al., 1991; Harman *et al.*, 2004). There are several mechanisms of genus *Trichoderma*, through which they compete with phytopathogen. Among them, the vital ones are grounded into triocategories: (i) Straight struggle for niche or supplements (Elad and Baker 1985; Elad and Chet 1987; Chet and Ibar 1994; Belanger *et al.*, 1995) (ii) Production of volatile or non-volatile antibiotic metabolites (Sid 2000; SIAP 2016) and (iii) mechanism of direct parasitism by few species of *Trichoderma* spp. (Yedidia *et al.*, 1999; Ezziyyani 2004). There are many reports which explained the use of different fungal biocontrol agents and VAM fungi to control of numerous root-borne diseases caused by *Fusarium* species (Dehne and Shoenbeck 1979).

## A. The Host Crop

Tomato (*Solanum lycopersicum*) is a flowering plant which belongs to nightshade family (Solanaceae). It is extensively grown for its palatable fruits. Tomatoes serve as rich source of vitamin C and lycopene and for its high nutritional values, they are labelled as a vegetable. The fruits are consumed in different ways like raw form as salads. Cooked vegetable is made and also utilised as an ingredient to make pickle and cook different dishes. Moreover, by-products of huge part of world's tomato crop are prepared like ketchup, puree, products, tomato juice, paste and dehydrated pulp. Canned tomatoes and "sun-dried" tomatoes are also available. Requirement of this plant requires generally warm weather and considerable sunlight. During cool climates, it is cultivated essentially in hothouses. To retain tomato stems and fruits off the ground, they are generally staked, tied, or caged and to escape blossomend rot and fruits cracking, constant irrigating is required (Adam *et al.*, 2019).

#### B. Constraints in Tomato Production

As it is a central crop cultivated worldwide (Fig. 1 and 2), there are numerous limitations which leads to huge loss in yield and seed production. The main reason of crop losses in worldwide is diseases of plants caused by fungi, bacteria, viruses or nematodes. Different pest and disease attacking tomato plants like Fusarium wilt, early blight, bacterial wilt, mosaicvirus, tomato hornworms and nematodes etc. In addition to pest's and pathogen's losses, post-harvest conditions also restrict the overall production of the crop. In the international markets, the food security norms have been strengthened and mycotoxins causing contamination of food have been acknowledged unfit for intake of human (WHO, 2002).

#### C. Wilt Disease of Tomato

#### Symptoms

Wilt of tomato is caused by *Fusarium oxysporum* f. sp. *lycopersici* (FOL) which is considered a major tomato disease (Borisade *et al.*, 2017). Epidermis of root is starting point, where the pathogen comes with contact of host. Pathogen penetrates epidermis and spreads to vascular tissue gradually. It occupies plant's xylem vessels and causes clogging of vessels resulting in acute stress of water, thus wilt like symptoms is ultimately observed (Singh *et al.*, 2017). Visually and morphologically, this disease is recognized by yellow

coloured leaves on partial or complete wilted plants. Fusarium movement in host is a very compound phenomenon, and the successive advances associated with its infection progression (Di *et al.*, 2016).

#### Causal Organism

FOL has three identified races (Races 1, 2 and 3) which are distinct by their principle resistance genes. Availability of races 1 and 2 has been reported from all the parts of world cultivating tomato, however presence of race 3 is stated in nations like Georgia, Mexico and California etc. In the whole, world maximum cultivated commercial varieties of tomato are found to be resistant against races 1 and 2, but against race 3 only few varieties show resistant (Biju et al., 2017). Small range transmission of FOL occurs primarily through water system and ranch hardware's which are contaminated by pathogen, whereas infected transplants, soils etc. play a major role in long distances transmission (Agrios, 2005). It has been reported that the fungus generally sustains indefinitely in an area, if once gets contaminated with FOL (Animashaun et al., 2017; Prihatna et al., 2018).

#### **Epidemiology and Losses**

The global dispersal of FOL is identified as cosmopolitan and surviving as soil saprophyte. This pathogen is extensively known among varied fungi and in agricultural soil, it is regarded as a prevalent fungus. This pathogen has wide dispersal area due to presence of the diverse formae speciales. Both in greenhouse and field conditions the influence of disease in tomato plant is at 28°C (Bawa 2016; Debbi *et al.*, 2018). Thus, this ailment causes huge loss in production of fruit i.e. nearly 60-70 percent along with dry wilted crops (Singh *et al.*, 2015).

# Infection Stages and Disease Cycle

Being a soil-born pathogen, FOL own characteristic to easily and broadly survive as dormant structure is known as chlamydospores in the soil and its germination is triggered by presence of host root. After the germination, infection hyphae are form which stick to root surface and gradually penetrate.







Fig. 2. Production (T) and percent share of tomato in top five chilli producing states in the India (NHB, 2017-18).

Invasion of mycelium occurs intercellularly in the cortical cells of root and eventually enters vascular framework via xylem pits. In this way, this pathogen exhibits an exclusive infection pathway, where initially it colonizes entire xylem vessels and then, further quickly colonizes the host completely. Microconidia are produced by fungus itself in the xylem vessels and after detachment, these are transported through sap stream to up part of plants. This causes mycelial penetration of the upper vessels as germination of microconidia occurs in that part. Vessel blockage due to accumulation of fungal hyphae, secretion of toxins and formation of gum, gel and tylose lead to the characteristic wilt symptoms appearance. Throughout this time, wilt pathogen restrictes in the xylem vessels, spread through parenchymatous tissue and reproduction occurs forming spores on plant surfaces like leaf, steam etc. (McGovern 2015; Joshi 2018).

## D. Disease Management Microbial pesticides

Pseudomonas fluorescens PF27 and P. putida PP15 both have proved their capability to defend plant against wilt disease.PF15 showed antagonistic activity and reduced of pathogen growth by 47% in vitro condition, mean while in PP27 showed comparatively less inhibition of mycelium i.e. 10 percent. A trial was performed in in-situ condition for understanding induced systemic resistance (ISR) for which they took split-root design and to gauge antagonistic activity and ISR both, split-root design was not used. Commencement of symptoms was deferred and lowering of kinetics in comparison to pathogen control was observed due to Fluorescent Pseudomonas. The severity of the disease was reduced by 37-72% based on McKinney's index, and incidence level by 7-36% (Lamia et al., 2017).

Hence, an experiment was conducted to estimate the impact of biocontrol agents with *Glomus mossae*, *G. fasciculatium, Trichoderma viride* and *T. Harzianum* to control wilt disease of tomato. Substantial drop in severity of wilt disease, improvement in chlorophyll

content and plant growth parameters were recorded due to use of biocontrol agent's various portion. With increment of biocontrol agent's dosage, there was additionally increase in growth attributes. It was observed that higher dosage of bioagent led to maximum development and increase in content of chlorophyll content in each pot following less dosages of treatments in comparison to control plant where the least plant development was detected (Wani and Mir 2009).

An experiment was performed where they explored capability of Trichoderma's ten isolates to protect tomato plant against Fusarium wilt pathogen added to it. They also observed effect of these isolates in the presence and absence of the pathogen on the growth tomato plant. Lincoln Bio-Protection Research Centre Culture Collection was used to get the isolates and seed raising mix (0.5% w/w) was inoculated by them to raise two glasshouse experiments. Tomato Fusarium wilt incidence was reduced significantly (P < 0.05) up to 69% by two Trichoderma isolates. Some plants only showed vascular discoloration. In the presence of the pathogen, an isolate increased the growth of plant by >50 percent, whereas there was no enhancement in the plant growth parameters by Trichoderma isolates, when pathogen was absent. (Ghazalibiglar et al., 2016)

In another research, use of rhizobacteria for the management of Fusarium wilt disease in tomato was studied. They characterized their antagonistic activity against F. oxysporum. They took ten rhizobacterial isolates and among them, the highest inhibition of fungal growth (31%) was shown by Bacillus amyloliquefaciens CS-1 followed by В. amyloliquefaciens PCfS which inhibited by 28% percent. Characterization for their useful qualities discovered that every isolate showed positive relation towards root colonization and variable results were found for solubilization of phosphate, Indole acetic acid production. production of antifungal secondary siderophore formation, secretion of metabolite. Hydrogen cyanide and disrupting enzymes formation salikechitinase etc. During greenhouse study, Fusarium wilt incidence was significantly suppressed by *Ochrobactrum intermedium* TRB-1, *B. amyloliquefaciens* PCfS *and B. amyloliquefaciens* CS - 1, and treated seedlings also showed enhanced the vigor index (VI) in comparison to untreated one (Gowtham *et al.*, 2016).

Under pot condition, the biocontrol ability contrary to Fusarium wilt by Funneliformis mosseae, Acaulospora laevis (arbuscular mycorrhizal fungi) and Trichoderma viride was evaluated. Results displayed considerable upsurge in growth of plant by entire bioagents. When F. mosseae, A. laevis and T. viride inoculated collectively, increased the height of plant, fresh weight of shoot, dry weight of root and leaves branches counts of each plant although F. mosseae and T. viride combined application augmented other growth parameters such as dry weight of shoot, fresh weight of root, root length and leaf area. On single inoculation of AMF, colonisation and spore number of AM was observed maximum, which diminishes on adding T. viride. Nevertheless, no effect was observed on the biocontrol efficacy of bioagents of this reduction. Pathogen infection caused considerable reduction in total amount of chlorophyll. photosynthesis, and nutrients. This effect was nullified by application of bioagent and leads to a notable rise in the nitrogen and phosphorus contents of the plant. Between the two AMF, for better tomato strain was F. mosseae, as it was more effective in comparison to A. laevis. The highest drop in incidence and severity of disease was documented in collective application of T. viride, F. mosseae and A. laevis. However, maximum disease incidence was observed in control plants without any bioagent. This study explains that, F. mosseae soil inoculation and application of T. viride conidial suspension in root former to transplantation causes improved immunity and subsistence capacity in seedlings of tomato against wilt disease (Tanwar et al., 2013).

In another experiment, in the tomato plant (Solanum *lycopersicum*) the potential of *T. virens* (TriV\_JSB100) using spores for treatment against F. oxysporum f. sp. lycopersici was studied and found its participation in the regulation and activation of the defence responses. Barley seeds(BGS) used as substrate for growth of Trichoderma spore or culture filtrate (CF) free of cell (TriV\_JSB100) was used for priming of tomato seeds, these fourteen-day old tomato seedlings were further inoculated with pathogen (F. oxysporum f. sp. lycopersici). Priming of bioagent caused considerable reduction in disease incidence in tomato plants, but BGS treated plant showed more reduction in comparison to plants treated with CF. Moreover, for analysing signalling behaviour in this condition, salicylic acid (SA) and jasmonic acid (JA) impaired tomato lines were used as to examine BGS and CF tempted immunity. They found BGS treated mutant which lack JA (def1) plants are prone to pathogen, however, less disease incidence and in BGS-treated wild type (WT) plants higher JA level were found. CF treated SA-deficient mutant NahG plants were also observed to exhibit less SA and also prone to pathogen. CF treated wild type plant showed less disease incidence and significantly increased SA. PDF1 (JAresponsive defensin gene) and PR1a (SA-inducible pathogenesis- related protein 1 gene) were expressed in BGS treated Wild type plants and Wild type plants treated with CF, respectively. These led to a suggestion that induction in plants treated with BGS and CF of TriV\_JSB100 causes differentially signalling cascades by jasmonic acid and salicyclic acid for the activation of resistance in tomato plant against Fusarium wilt (Jogaiah *et al.*, 2018).

In vitro bioassay was done in Egypt for the assessment of antagonistic capability of seven isolates of Trichoderma spp. against F. oxysporum f. sp. lycopersici. Maximum inhibition percentage against the pathogen was shown by Trichoderma isolate (T7) followed by T3 isolate. Substantial decline in the disease severity was observed in plants treated with T3 and T7 isolate in comparison to the control treatment under greenhouse trials. In comparison to other isolates, the lowest disease severity was 24.8% which shown by T3 isolate and succeeded by 34.6% via T7 isolate. Realtime RT-PCR was used to quantify the expression of defense-related -1,3-glucanase gene for assessment of the accumulation kinetics of transcripts encoding PR proteins in the roots of tomato in control plant i.e inoculated with only pathogen, plant treated with T3 isolate and wilt pathogen, and plant treated withT7 isolate and wilt pathogen. Maximum expression of gene was reported in tomato plant treated with T3 and FOL in comparison to control. Using universal primer internal transcribed spacers (ITS1 and ITS4), these two species (T3 and T7) of Trichoderma showing antagonistic activity were characterized which evidenced that used isolates were not the same i.e. T. atroviride and T. longibrachiatum and belonged to different species (Sallam et al., 2019).

Host could acquire resistance against phytopathogens either by direct antagonism or by activating induced resistance via endophytes of root. Induced systemic resistance (ISR) basically relies on signaling pathways of jasmonic acid (JA)/ethylene (ET), but salicylic acid (SA)-dependent signaling pathways could also trigger it. An experiment was conducted to explore the involvement of Jasmonic acid, Saliscylic acid and Ethylene in endophyte-mediated resistance (EMR) conferred by Fusarium endophyte Fo47 against wilt of tomato. The investigations incorporate impaired tomato plants which deduced accumulation of SA (NahG), JA formation (def1) or formation of ET (ACD) and detecting (Nr). Pattern of Fo47colonization in stems was indistinguishable among wild type plants and of hormone mutants. Unexpectedly, JA, ET, or SA impaired lines does not show any compromise in EMR, which showed EMR not dependent on JA, ET, or SA signalling. Thus, it explained difference between classical Induced Systemic Resistance (ISR and induced resistance conferred against tomato wilt disease caused by F. oxysporum f.sp. lycopersici). This experiment concluded that management of Fusarium wilt is self-regulating and don't rely on basic defense mechanisms (Constantin et al., 2019).

To understand induction of phenolic compounds accumulation and defense enzymes in *F. oxysporum* f.

sp. lycopersici- infected tomato plants under treatment of salicylic acid (SA) and a biocontrol agent(T.harzianum), an experiment was conducted. In each treatment (F. oxysporum, F. oxysporum + TH, F. oxysporum + SA and F. oxysporum + TH + SA), accumulation of phenolic compounds was increased in comparison to control healthy plants. In the plants, the highest accumulation was recorded under the combined application of F. oxysporum + TH + SA. The accumulation of peroxidase and polyphenol oxidase in plants was raised due to application of salicylic acid at varied concentrations i.e. 0.5, 1.0, 1.5, 2.0, 2.5 and 3.0 mM, though at amount,1.5 mM concentration of SA was found the most effective as it showed the highest actions of peroxidase and polyphenol oxidase enzymes on  $28^{\text{th}}$  day after the combined application of F. oxysporum and salicyclic acid. The same pattern of enhancement in the activities of both enzymes was obtained in Fusarium-infected tomato plants, treated with 1.5 mM of salicyclic acid and T. harzianum (Ojha and Chatterjee 2012).

Biological control of wilt of tomato has been also explored where biofortified vermicompost containing biological control agents (*T. harzianum*, *P. fluorescens* and *B. subtilis*) has been used. Adversarial effect of chosen microorganisms against FOL has been reported in *in-vitro* test. In intended treatments, parameters of plant growth, degrees of various antioxidants and disease occurrence were documented at various time spans which described huge decrease in occurrence of disease, improvement in growth of plant, increase in production along with increased production of antioxidants in vermicompost - applied tomato plants in comparison to untreated plants. Host plant treated with vermicompost biofortified with *T. harzianum* showed utmost improvement (Basco *et al.*, 2017).

## CONCLUSION

For the management of Fusarium wilt disease in tomato, many synthetic chemical pesticides are in use. These pesticides are affecting our natural environment, while combating Fusarium wilt disease. To reduce the hazardous effect of pesticide and meet the food demand of increasing world population, researchers investigated and developed different alternative ways for management of diseases. One of them is the use of natural biocontrol agent which has been proved economic and ecological alternative method to manage different plant diseases in effective manner. Many investigations showed a potential role of biocontrol agent against plant diseases management. However, research and knowledge gap of plant microbe interactions, especially for the plant-biotic stressbiocontrol agents in rhizosphere region. Improvement in knowledge about rhizosphere ecology interactions and distribution of antagonistic and pathogenic microorganisms can help in enhancing the efficacy of bio-agents against plant diseases.

## FUTURE SCOPE

All numerous techniques to control fungal diseases has different mode of action. Thus a comprehensive ecofriendly method including considerate pathogen virulence and genetic diversity, host resistance, and plant-pathogen interactions should be taken into consideration to help a build integrated management solutions. Moreover for advancement of microbial pesticides control method emphasis should be given to determine and improve their efficiency, effectiveness, and long-term viability in the field, not merely in the controlled conditions. Training farmers on the proper application of these microbial pesticides and their incorporation into other tactics for a healthier and nontoxic produce should be given a priority.

**Conflict of Interest.** The authors declare no conflict of interest.

Abbreviations. PGPR: Plant Growth Promoting Rhizobacteria, ISR: Induced Systemic Resistance, AMF: Arbuscular Mycorrhizal Fungi, SA: Salicylic Acid, JA: Jasmonic Acid, CF: Culture Filtrate, ET: Ethylene, EMR: Endophyte Mediated Resistance, WHO: World Health Organization, FOL: *Fusarium oxysporum* f.sp. *lycopersici*.

## Authors' contributions

All authors participated in the development and implementation of the reviewing plan and subsequently written it. The first author RB discussed the different parts of the article with GK and finalized the manuscript. All authors have read and approved the final manuscript.

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## REFERENCES

- Abdallah, R.A.B., Mokni-Tlili, S., Nefzi, A., Jabnoun-Khiareddine. H. and Daami-Remadi, M. (2016). Biocontrol of Fusarium wilt and growth promotion of tomato plants using endophytic bacteria isolated from *Nicotiana glauca* organs. *Biological Control*, 97: 80-88.
- Adam, A., Patricia, B., Brian, D., Alison, E., Erik, G., Amy, M., Melissa, P., John, R., Michael, R., Kara, R., Amy, T., Jeff, W., Adam, Z. and Alicja, Z. (2019). Tomato. Encyclopaedia Britannica. Inc. https://www.britannica.com/plant/tomato
- Agrios, G. N. (2005). Plant diseases caused by fungi. *Plant Pathology*, *4*: 385-614.
- Animashaun, B.O., Popoola, A.R., Enikuomehin, O.A., Aiyelaagbe, I.O.O. and Imonmion, J.E. (2017). Induced resistance to Fusarium wilt (*Fusarium* oxysporum) in tomato using plant growth activator, acibenzolar-S-methyl. Nigerian Journal of Biotechnology, 32: 83-90.
- Arenas, O. R., Olguín, J. F. L., Ramón, D. J., Sangerman-Jarquín, D. M., Lezama, C. P., Morales, P. S. and Lara, M. H. (2018). Biological control of *Fusarium* oxysporum in tomato seedling production with Mexican strains of *Trichoderma*. Fusarium—plant diseases, pathogen diversity, genetic diversity, resistance and molecular markers, 155-168.
- Awad, H.M., El-Enshasy, H.A., Hanapi, S.Z., Hamed, E.R. and Rosidi, B. (2014). A new chitinase-producer strain

*Streptomyces glauciniger* WICC-A03: isolation and identification as a biocontrol agent for plants phytopathogenic fungi. *Natural Product Research*, 28: 2273-2277.

- Aydi-Ben-Abdallah, R., Jabnoun-Khiareddine, H. and Daami-Remadi, M. (2020). Fusarium wilt biocontrol and tomato growth stimulation, using endophytic bacteria naturally associated with *Solanum sodomaeum* and *S. bonariense* plants. *Egyptian Journal of Biological Pest Control*, 30: 1-13.
- Basco, M.J., Bisen, K., Keswani, C. and Singh, H.B. (2017). Biological management of Fusarium wilt of tomato using biofortified vermicompost. *Mycosphere*, 8: 467-483.
- Bawa, I. (2016). Management strategies of Fusarium wilt disease of tomato incited by *Fusarium oxysporum* f. sp. lycopersici (Sacc.) A Review. International Journal of Advanced Academic Research, 2: 32-42.
- Belanger, M.J., Miller, J.R. and Boyer, M.G. (1995). Comparative relationships between some red edge parameters and seasonal leaf chlorophyll concentrations. *Canadian Journal of Remote Sensing*, 21: 16-21.
- Benhamou, N., Kloepper, J.W. and Tuzun, S. (1998). Induction of resistance against Fusarium wilt of tomato by combination of chitosan with an endophytic bacterial strain: ultrastructure and cytochemistry of the host response. *Planta*, 204: 153-168.
- Biju, V.C., Fokkens, L., Houterman, P.M., Rep, M. and Cornelissen B.J. (2017). Multiple evolutionary trajectories have led to the emergence of races in *Fusarium oxysporum* f. sp. lycopersici. Applied and Environmental Microbiology, 83.
- Borisade, O.A., Uwaidem, Y.I. and Salami, A.E. (2017). Preliminary report on *Fusarium oxysporum* f. sp. *lycopersici* (Sensu lato) from some tomato producing agroecological areas in Southwestern Nigeria and susceptibility of F1-resistant tomato hybrid (F1-Lindo) to infection. *Annual Research & Review in Biology*, 1-9.
- Boukerma, L., Benchabane, M., Charif, A. and Khélif, L. (2017). Activity of plant growth promoting rhizobacteria (PGPRs) in the biocontrol of tomato Fusarium wilt. *Plant Protection Science*, 53: 78-84.
- Candela, M.E., Alcazár, M.D., Espín, A., Egea-Gilabert, C. and Almela, L. (1995). Soluble phenolic acids in *Capsicum annuum* stems infected with *Phytophthora capsici*. *Plant Pathology*, 44: 116-123.
- Chen, C., Belanger, R.R., Benhamou, N. and Paulitz, T.C. (2000). Defense enzymes induced in cucumber roots by treatment with plant growth-promoting rhizobacteria (PGPR) and *Pythium aphanidermatum*. *Physiological and Molecular Plant Pathology*, 56: 13-23.
- Chet, I., Benhamou, N. and Haran, S. (1998). Mycoparasitism and lytic enzymes. In: Harman GE, Kubicek CP, editors. *Trichoderma* and *Gliocladium*. Vol. 2. *London, UK: Taylor and Francis*, 153-172.
- Chet, I. and Inbar, J. (1994). Biological control of fungal pathogens. Applied Biochemistry and Biotechnology, 48: 37-43.
- Constantin, M.E., de Lamo, F.J., Vlieger, B.V., Rep, M. and Takken, F.L. (2019). Endophyte-mediated resistance in tomato to *Fusarium oxysporum* is independent of ET, JA, and SA. *Frontiers in Plant Science*, 10: 979.
- Cook, R.J. and Baker, K.F. (1989). The Nature and Practice of Biological Control of Plant Pathogens. *American Phytopathological Society*, 589.
- Debbi, A., Boureghda, H., Monte, E. and Hermosa, R. (2018). Distribution and genetic variability of *Fusarium*

oxysporum associated with tomato diseases in Algeria and a biocontrol strategy with indigenous Trichoderma spp. Frontiersin Microbiology, 9: 282.

- Dehne, H.W. and Schoenbeck, F. (1979). "The influence of endotrophic mycorrizae on plant disease. I. Colonization of tomato plants by *Fusarium* oxysporum. 105-105.
- Di, X., Takken, F.L. and Tintor, N. (2016). How phytohormones shape interactions between plants and the soil-borne fungus *Fusarium oxysporum*. *Frontiersin Plant Science*, 7: 170.
- Elad, Y. and Baker, R. (1985). Influence of trace amounts of cations and siderophore-producing *Pseudomonads* on chlamydospore germination of *Fusarium oxysporum*. *Phytopathology*, 75: 1047-1052.
- Elad, Y. and Chet, I. (1987). The role of competition for nutrients in biocontrol of *Pythium* damping off by bacteria. *Phytopathology*, 77: 190-195.
- Ezziyyani, M.E., Pérez, S.C., Requena, M.E., Rubio, L. and Candela, C.M.E. (2004). Biocontrol por *Streptomyces rochei* –Ziyani de la podredumbre del pimiento (*Capsicum annuum* L.) causada por *Phytophthora capsici. Anales de Biologia*, 26: 69-78.
- Ghazalibiglar, H., Kandula, D.R. and Hampton, J.G. (2016). Biological control of Fusarium wilt of tomato by *Trichoderma* isolates. *New Zealand Plant Protection*, 69:57-63.
- Ghisalberti, E.L. and Sivasithamparam, K. (1991). Antifungal antibiotics produced by *Trichoderma* spp. Soil Biology and Biochemistry, 23:1011-1020.
- Giovannucci, E. (1999). Tomatoes, tomato-based products, lycopene, and cancer: review of the epidemiologic literature. *Journal of The National Cancer Institute*, 91: 317-331.
- Giovannucci, E., Rimm, E.B., Liu, Y., Stampfer, M.J. and Willett, W.C. (2002). A prospective study of tomato products, lycopene, and prostate cancer risk. *Journal* of *The National Cancer Institute*, 94: 391-398.
- Glick, B.R. and Bashan, Y. (1997). Genetic manipulation of plant growth-promoting bacteria to enhance biocontrol of fungal phytopathogens. *Biotechnology Advance*,15: 353-378.
- Gowtham, H.G., Hariprasad, P., Nayak, S.C. and Niranjana, S.R. (2016). Application of rhizobacteria antagonistic to *Fusarium oxysporum* f. sp. *lycopersici* for the management of Fusarium wilt in tomato. *Rhizosphere*, 2: 72-74.
- Hamed, E.R., Awad, H.M., Ghazi, E.A., El-Gamal, N.G.and Shehata, H.S. (2015). *Trichoderma asperellum* isolated from salinity soil using rice straw waste as biocontrol agent for cowpea plant pathogens. *Journal* of Applied Pharmaceutical Science, 5: 091-098.
- Harman, G.E., Howell, C.R., Viterbo, A., Chet, I. and Lorito, M. (2004). *Trichoderma* species opportunistic, avirulent plant symbionts. *Nature Reviews Microbiology*, 2: 43-56.
- Howell, C.R. and Stipanovic, R.D. (1995). Mechanisms in the biocontrol of *Rhizoctonia solani* induced cotton seedling disease by *Gliocladium virens: Antibiosis*. *Phytopathology*, 85: 469-472.
- Jogaiah, S., Abdelrahman, M., Tran, L.S.P. and Ito, S.I. (2018). Different mechanisms of *Trichoderma virens*mediated resistance in tomato against Fusarium wilt involve the jasmonic and salicylic acid pathways. *Molecular Plant Pathology*, 19:870-882.
- Joshi, R. (2018). A review of Fusarium oxysporum on its plant interaction and industrial use. Journal of Medicinal Plants Studies, 6: 112–115.
- Kloepper, J.W. and Beauchamp, C.J. (1992). A review of issues related to measuring colonization of plant roots

by bacteria. *Canadian Journal Microbiology*, 38:1219-1232.

- Kloepper, J.W., Schroth, M.N. and Miller, T.D. (1980). Effects of rhizosphere colonization by plant growth promoting rhizobacteria on potato plant development and yield. *Phytopathology*, 70: 1078-1082.
- Kouki, S., Saidi, N., Ben Rajeb, A., Brahmi, M., Bellila, A., Fumio, M. and Ouzari, H. (2012). Control of Fusarium wilt of tomato caused by *Fusarium oxysporum* f. sp. *radicis-lycopersici* using mixture of vegetable and *Posidonia oceanica* compost. *Applied and Environmental Soil Science*.
- Lamia, B., Messaoud, B., Ahmed, C. and Lakhdar, K. (2017). Activity of plant growth promoting rhizobacteria (PGPRs) in the biocontrol of tomato Fusarium wilt. *Plant Protection Science*, 53: 78-84.
- Liu, L., Kloepper, J.W. and Tuzun, S. (1995). Induction of systemic resistance in cucumber against, Fusarium wilt by plant growth promoting rhizobacteria. *Phytopathology*, 85: 695–698.
- Loganathan, M., Garg, R., Venkataravanappa, V., Saha, S. and Rai, A.B. (2014). Plant growth promoting rhizobacteria (PGPR) induces resistance against Fusarium wilt and improves lycopene content and texture in tomato. *African Journal of Microbiological Research*, *11*: 1105-1111.
- Loganathan, M., Rai, A.B., Ramesh, R., Sharma, B.K., Rai, R.K. and Rai, M. (2009). Vascular wilt disease-a menace in vegetable crops. *Vegetable Science*, 36: 1-13.
- McGovern, R.J. (2015). Management of tomato diseases caused by *Fusarium oxysporum*. Crop Protection, 73: 78-92.
- Mohammed, B.L. and Toama, F.N. (2019). Biological control of Fusarium wilt in tomato by endophytic rhizobactria. *Energy Procedia*, 157: 171-179.
- M'piga, P., Belanger, R.R., Paulitz, T.C. and Benhamou, N. (1997). Increased resistance to *Fusarium oxysporum* f. sp. *radicis-lycopersici* in tomato plants treated with the endophytic bacterium *Pseudomonas fluorescens* strain 63-28. *Physiological and Molecular Plant Pathology*, 50: 301-320.
- Ojha, S. and Chatterjee, N. (2012). Induction of resistance in tomato plants against *Fusarium oxysporum* f. sp. *lycopersici* mediated through salicylic acid and *Trichoderma harzianum. Journal of Plant Protection Research*, 52: 220-225.
- Prihatna, C., Barbetti, M.J. and Barker, S.J. (2018). A novel tomato Fusarium wilt tolerance gene. *Frontier in Microbiology*, 9: 1226.
- Sallam, N.M., Eraky, A.M. and Sallam, A. (2019). Effect of *Trichoderma* spp. on Fusarium wilt disease of tomato. *Molecular Biology Report*, 46: 4463-4470.
- Sandoval, V.M.C. and ZMCI, N. (2011) Producción de conidios de *Trichoderma harzianum rifai* en dos medios de multiplicación. *Fitosanidad*, 15: 215-221.
- Sangeetha, G., Thangavelu, R., Rani, S. U., Muthukumar, A., and Udayakumar, R. (2010). Induction of systemic resistance by mixtures of antagonist bacteria for the management of crown rot complex on banana. Acta Physiologiae Plantarum, 32: 1177-1187.

Saravanakumar, D., Vijayakumar, C., Kumar, N. and Samiyappan, R. (2007). PGPR-induced defense responses in the tea plant against blister blight disease. *Crop Protection*, 26: 556-565.

- SIAP (Servicio de Información Agroalimentaria y Pesquera de la SAGARPA) (2016). Rendimientos de granos por Estados y años. https:// www.gob.mx/siap/acciones-yprogramas/produccion-agricola-33119
- Ahmed, A. S., Sánchez, C. P., and Candela, M. E. (2000). Evaluation of induction of systemic resistance in pepper plants (*Capsicum annuum*) to *Phytophthora capsici* using *Trichoderma harzianum* and its relation with capsidiol accumulation. *European Journal of Plant Pathology*, 106: 817-824.
- Singh, R., Biswas, S.K., Nagar, D., Singh, J., Singh, M., Mishra, Y.K. (2015). Sustainable integrated approach for management of Fusarium wilt of tomato caused by *Fusarium oxysporum* f. sp. lycopersici (Sacc.) Synder and Hansen. Sustainable Agriculture Research, 4: 138-147.
- Singh, V.K., Singh, H.B. and Upadhyay, R.S. (2017). Role of fusaric acid in the development of 'Fusarium wilt' symptoms in tomato: Physiological, biochemical and proteomic perspectives. *Plant Physiology and Biochemistry*, 118: 320-332.
- Tanwar, A., Aggarwal, A. and Panwar, V. (2013). Arbuscular mycorrhizal fungi and *Trichoderma viride* mediated Fusarium wilt control in tomato. *Biocontrol Science* and *Technology*, 23: 485-498.
- Upadhyay, A.K., Bandi, S.R. and Peddaguad, D.M. (2021). BioControl Agents - Antagonistic Magicians against Soil Borne Pathogens: A Review. *Biological Forum – An International Journal*, 13: 232-242.
- Vargas-Inciarte, L., Fuenmayor-Arrieta, Y., Luzardo-Méndez, M., Costa-Jardin, M.D., Vera, A., Carmona, D. and San-Blas, E. (2019). Use of different *Trichoderma* species in cherry type tomatoes (*Solanum lycopersicum* L.) against *Fusarium oxysporum* wilt in tropical greenhouses. *Agronomía Costarricense*, 43: 85-100.
- Verma, M., Brar, S.K., Tyagi, R.D., Surampalli, R.Y. and Valero, J. R. (2007). Antagonistic fungi, *Trichoderma* spp.: panoply of biological control. *Biochemical Engineering Journal*, 37: 1-20.
- Wang, C., Knill, E., Glick, B.R. and Défago, G. (2000). Effect of transferring 1-aminocyclopropane-1-carboxylic acid (ACC) deaminase genes into *Pseudomonas fluorescens* strain CHA0 and its *gacA* derivative CHA96 on their growth-promoting and diseasesuppressive capacities. *Canadian Journal of Microbiology*, 46: 898-907.
- Wani, A.H. and Mir, R.A. (2009). Biological control of *Fusarium oxysporum* f. sp. lycopercici on tomato with fungal antagonists. *Journal of Biological Control*, 23: 169-173.
- WHO (2002). Evaluation of Certain Mycotoxins in Food, Fifty-Sixth Report of the Joint FAO/WHO Expert Committee on Food Additives. WHO Technical Report Series 906, Geneva. Available online at: http://www.who.int/iris/handle/10665/42448
- Yedidia, I., Benhamou, N. and Chet, I. (1999). Induction of defense responses in cucumber plants (*Cucumis* sativus L.) by the biocontrol agent *Trichoderma* harzianum. Applied and Environmental Microbiology, 65: 1061-1070.

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